**COMPARISON OF PHENOLIC, FLAVONOID, AND TANNIN CONTENTS FROM ETHANOL EXTRACT OF KRATOM STEM (*Mitragyna speciosa* Korth.) AND SENGGANI FLOWER (*Melastoma malabathrium* L.)**

**H.N. Hanifah1 , G. Hadisoebroto2 , L. Dewi3**

Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Al-Ghifari University,

Cisaranten Kulon Street, Bandung, Indonesia

**\*Correspondence Author: hesty.nuur@gmail.com**

**Abstract.** Indonesia is a country known for its useful natural resources, especially for a health maintenance and treatment of diseases. Some of the plants that can be used as traditional medicine are Kratom (*Mitragyna speciosa* korth) and Senggani (*Melastoma malabathrium* L.). Kratom plants contain alkaloid compounds, flavonoids, tannins, phenols and triterpenoids, while Senggani flowers contain saponins and tannins. Phenolic that spread throughout the plant can be used as a natural antioxidants. Flavonoids, that are included in phenolic compounds group, functioned as free radical scavengers, inhibitors of hydrolysis and oxidative enzymes, and anti-inflammatory. Tannins are reported to have antioxidant, antibacterial, antiviral, anti-inflammatory, allergy and anti-cancer activities. The purpose of this research was to determine the total phenolic content, total flavonoids content, and total tannins content in the ethanol extract of kratom stems and flowers using the UV-Vis spectrophotometric method. The comparator used in determining the phenolic content was gallic acid, for the flavonoid content was quercetin, and for the tannin content was tannic acid. The results of this study showed that the total phenolic content in the kratom stem extract was 23.59 mg/gram of extract, while in the senggani flower was 43.97 mg/gram of extract. The total flavonoid content in the kratom stem extract was 10.65 mg/gram of extract, while the senggani flower was 11.26 mg/gram of extract. Lastly, the total tannin content in the kratom stem extract was 17.99 mg/gram of extract, while in the senggani flower it was 13.74 mg/g extract. According to the results, the phenolic and flavonoid levels of senggani flower extract were greater than that of kratom stems, while the tannin levels of kratom stem extract were greater than that of senggani flowers.

1. **Introduction**

Indonesia is a country known for its good usage of nature, particularly as a health maintenance and treatment of diseases (Sahara et al., 2019). Some of the plants that can be used as traditional medicine are Kratom (*Mitragyna speciosa* korth) and Senggani Flower (*Melastoma malabathrium* L.). Both of those plants were native to West Kalimantan Province. Kratom plant (*Mitragyna speciosa* korth) was a plant from Southeast Asia, where the plant has been used as a medicinal plant since ancient times. The indigenous people used this plant as medicine to restore their strength (Ridayani, 2013). Senggani Flower (*Melastoma malabathrium* L) was known as natural dye. This plant contains saponins, flavonoids, and tannins. These compounds are effective as fever relievers (antipyretics), pain relievers (analgesics), urinary laxatives (diuretics), eliminate swelling, improve blood flow, and stop bleeding (hemostasis) (Sahara et al., 2019). Phenolic compounds are natural compounds that have wide range of uses. The phenol group consists of several compounds, including flavonoids and tannins. Its ability as a biologically active compound provides a big role for human. One of them is for the treatment of degenerative diseases. Flavonoid compounds are found in almost all parts of the plant, including fruit, roots, leaves, and stems. Flavonoids and tannins are reported to have antioxidant, antibacterial, antiviral, anti-inflammatory, anti-allergy, and anti-cancer activity (Alfian et al., 2012). This prompted researchers to analyze the total phenolic content, total flavonoid content, and total tannin content from kratom stem extract and senggani flower so that they could be used as herbal medicine. The research used a UV-Vis spectrophotometric instrument to measure the contents. The method used to analyze the content in the plant was to compare it with standard solutions. The standard solution for flavonoid was quercetin, for phenolic was gallic acid, and for tannin was tannic acid.

1. **MATERIAL AND METHODS**

**2.1 Equipments**

The equipments used for this research were the Shimadzu UV-Vis spectrophotometer, rotary vacuum evaporator, moisture balance, oven, and other tools in the natural product laboratory.

### Materials

The materials used for this research were kratom stem, Senggani flower, ethanol 96%, aquades, aluminium chloride (AlCl3) 10%, FeCl3 1%, quercetin, gallic acid, tannic acid, potassium acetate 1 M, concentrated sulfuric acid (H2SO4), Folin-Ciocalteau reagent, magnesium-chloric acid (Mg-HCl), and Na2CO3 7%.

#### Plant Determination

Plant determination was carried out at the Faculty of Agriculture, Laboratory of Land Quality and Health, Tanjungpura University (UNTAN), Pontianak, West Kalimantan.

**2.4 Water Content**

#### The water content in simplicia was determined by using Moisture Balance. A total of 2 g of sample was put into the Moisture Balance that had been prepared at 100°C for 10 minutes. The levels listed on the Moisture Balance were then recorded (Wiendarlina et al., 2018).

#### Maceration Extraction

The kratom stem extract and senggani flower extract were made by using the maceration method. 200 grams of simplicia each was put into a maceration vessel, soaked using 96% ethanol solvent until all samples were immersed and stirred for 30 minutes until completely mixed, after that, it was left for 24 hours and then filtered. The process was carried out for 3 x 24 hours . The maceration results were filtered and ethanol filtrate was obtained. The ethanol filtrate was then concentrated at a temperature of 30-45ºC by using rotary vacuum evaporator to obtain a thick ethanol extract of kratom stems and ethanol extract of senggani flowers.

. The yield was determined by using equation:

|  |
| --- |
| Yield = |

#### Phytochemical Screening

Each medicinal plant contains a variety of organic compounds that are formed and contained in these plants. The content of active compounds contained in plants can be determined by separation, purification, and phytochemical screening. Phytochemical screening includes:flavonoid test, phenolic test, tannin test.

#### Determination of the Total Phenolic Content of Kratom Stem Extract

* + 1. *Determination of Maximum Wavelength*

Prepare a total of 10 mg gallic acid, put into a 10 mL volumetric flask and dissolved in 10 mL of distilled water. Then 1 mL pipette was added and 0.4 Folin-Ciocalteu reagent was shaken, left for 3 minutes. Then add 4.0 mL of 7% Na2CO3 solution, shake until homogeneous and add distilled water to 10 mL, stand for 40 minutes (Ahmad et al, 2015). Then the absorbance is measured at a wavelength of 600-850 nm (Apsari et al., 2011).

* + 1. **Preparation of Gallic Acid Curves with Folin-Ciocalteu Reagent**

10 mg of gallic acid was diluted in 10 mL of distilled water on a volumetric flask. From this solution, 0.2; 0.4; 0.6; 0.8; and 1 mL were taken and mixed with distilled water up to 10 mL to make a standard concentration of 20 ppm, 40 ppm, 60 ppm, 80 ppm, and 100 ppm. 1 mL of each standard solution of gallic acid was mixed with 0.4 mL of Folin-Ciocalteu reagent. The mixture was shaken and left for 3 minutes and added with 4.0 mL of 7% Na2CO3 solution. The mixture was shaken until homogeneous and added with 10 mL of distilled water and then incubated for 40 minutes (Ahmad et al, 2015).

**2.7.3 Determination of Total Phenolic**

100 mg of ethanolic extract of kratom stem was dissolved in 100 mL of distilled water. 1 mLof samples was added with 0.4 mL of Folin-Ciocalteu reagent, and shaked the solution and left for 3 minutes. 4.0 mL of 7% Na2CO3 solution was added to the mixture and shaked the mixture until homogeneous. 10 mL of distilled water was added to the mixture and then incubated for 40 minutes (Ahmad et al., 2015). The absorbance of the extract solution was measured using UV-Vis spectrophotometry with the results of the obtained wavelength of gallic acid. The measurements were made three times. Data analysis was performed using the standard curve method, linear regression was made based on absorbance data and the concentration of gallic acid standard solution (Alfian et al., 2012). The same procedure was repeated for the ethanolic extract of senggani flowers.

**2.8 Determination of Total Flavonoid of Ethanolic Extract of Kratom Stem and Ethanol Extract of Senggani Flowers**

*2.8.1 Determination of Maximum Wavelength*

The maximum wavelength is determined based on the Ahmad et al., (2015) journal with a modification of 10 mg of quercetin inserted into a 10 ml volumetric flask and dissolved in 10 ml of 96% ethanol. 0.1 ml of the solution was added with 0.2 mL of aluminum chloride (AlCl3) 10%, and added with 0.2 ml of potassium acetate 1 M and then added with distilled water up to 10 ml. The absorbance was measured at a wavelength of 400 - 800 nm using UV-Vis spectrophotometry (Krisyanella et al., 2012).

*2.8.2 Preparation of Quercetin Standard Curves with Folin-Ciocalteu Reagent*

A standard quercetin curve was made based on the Ahmad et al., (2015) journal with a modified of 10 mg of quercetin was inserted into a 10 ml volumetric flask and dissolved in 10 ml of 96% ethanol for a concentration of 1000 ppm. 1 ml of the solution was dissolved in 10 ml of ethanol 96 % for a concentration of 100 ppm and then several standard concentrations of 20 ppm, 40 ppm, 60 ppm, 80 ppm were made. 1 mL of each standard solution of quercetin was added with 0.2 ml of 10% aluminum chloride (AlCl3), 0.2 ml of 1 M potassium acetate and added up to 10 ml with distilled water. The absorbance of all standards were measured at the quercetin wavelength using UV-Vis spectrophotometry that has been obtained.

* + 1. *Determination of Total Flavonoid*

Determination of total flavonoid by colorimetric method based on the Ahmad et al., (2015) journal with modification, 100 mg of kratom stem ethanol extract dissolved in 100 ml of distilled water. 0.1 ml of the samples was added with 0.2 ml of 10% AlCl3, 0.2 ml of 1 M potassium acetate and then added with distilled water up to 10 mL. The absorbance was measured at a maximum wavelength of quercetin using UV-Vis spectrophotometry. Measurements were made 3 times. Data analysis was performed using standard linear regression curve method based on absorbance data and concentration of quercetin standard solution. The same procedure was repeated for the ethanolic extract of senggani flowers.

**2.9 Determination of Total Tannin Levels of Ethanol Extract of Kratom Stem and Ethanol Extract of Senggani Flowers**

*2.9.1 Determination of Wavelength*

0.01 gram of tannic acid was dissolved in 10 mL of distilled water. 1 mL of tannic acid was put into a 10 mL volumetric flask and then 0.4 mL of Folin-Ciocalteu reagent was added. The mixture was shaken homogeneously and left for 3 minutes and then added with 1 mL of 7% Na2CO3  solution, shaken homogeneously and added with distilled water up to 10 mL and then incubated for 40 minutes (Mukhriani et al, 2014). The absorbance was masured at 400-800 nm using UV-Vis spectrophotometry (Kharismawati et al., 2009).

*2.9.2 Preparation of Tanic Acid Standard Curves with Folin-Ciocalteu Reagent*

0.01 grams of tannic acid was dissolved in 10 mL of distilled water for a concentration of 1000 ppm. Then several standard concentrations of 20 ppm, 40 ppm, 60 ppm, 80 ppm, 100 ppm were made. 1 mL of each concentration was put into a 10 mL volumetric flask and was added with 0.4 mL of the Folin-Ciocalteu reagent. The solution was shaken homogeneously and left for three minutes then added with 1 mL of 7% Na2CO3 solution, shaken until homogeneous and added with distilled water up to 10 mL and incubated for 40 minutes (Mukhriani et al., 2014). The absorbance was measured at a maximum wavelength of tannic acid using UV-Vis spectrophotometry, and then a calibration curve is made.

*2.9.3 Determination of Tannin Content*

0.1 gram of kratom stem ethanol extract was dissolved in 100 mL distilled water. 1 mL of the samples was added with 0.4 mL of Folin-Ciocalteu reagent, and shaken the solution homogeneously and left for three minutes and then added 1 mL of 7% Na2CO3 solution. The mixture was shaken homogeneously and added with distilled water up to 10 mL and then incubated for 40 minutes (Mukhriani et al., 2014).

**III. RESULTS AND DISCUSSION**

**3.1 Determination Results**

Determination is carried out to find out the correct identity of the plants. Based on the letter of determination, the plant used is true kratom (*Mitragyna speciosa Korth.).*

**3.2 Moisture Content**

Determination of water content was important to gave the limit amount of water contained in simplicia, because the high amount of water can became a growth medium for bacteria and fungi that can damaged the compounds which contained in simplicia. The result of the water content for kratom stems was 3.8%. The result of the water content test for senggani flowers was 1%, this result showed that the simplicia was qualified for the water content requirements becaused the maximum water content limit for stem simplicia was not more than 10% (Agoes., 2007)

**3.3 Results of Phytochemical Screening**

Phytochemical screening was aim to ensured the chemical compounds contained in the simplicia and to ensured that the extraction process in the concentration of the extract did not damage these compounds so it was carried out on the ethanolic extract of kratom stems and senggani flower extracts.

In test phenolic shows the results of the positive things is because the phenolic reacted with FeCl3  1 % form the color red, purple, blue, or black are concentrated because of FeCl 3 1% reacts with the group -OH aromatic ( Haryati *et al*., 2015). The flavonoid test showed results   Positive thing is because of the addition of powdered   magnesium  and acid chlorides in the test will lead to its reduced compounds   of flavonoids  exist that give rise to a reaction color of red , yellow , and orange   which  is characteristic of the   flavonoids (Robinson, 1995). The tannin test shows positive results this is due to the formation of compounds complex with the formation of the color green -black or blue dark because the tannins will form a compound complex with the ion Fe 3+ (Harbone, 1987).

**3.4 Results of Making Kratom Stem and Senggani Flowers Extract**

             The purpose of extraction process is to attract the desired compound by using a suitable solvent, in the present study the extraction is done by maceration method, the advantage of this method is that it can prevent damage to the compound having thermolabile properties. A total of 200 g of simplicia was macerated using 1 L of 96% ethanol for 3 days and the solvent was replaced every 24 hours. Concentration of the liquid extract using a *rotary vaporator* with a temperature of 30-45 C and then stored in a water bath until it becomes a thick extract, the randemen value of kratom stem extract obtained is 44.5% while the yield value of senggani flower extract obtained is 52.5 %.

**3.5 Result of Determination of Maximum Wavelength**

Determination of total phenolic content is determined by the standard curve method using a UV-Vis spectrophotometry, determination of the maximum wavelength is carried out to determine the wavelength that has the greatest absorbance.

Table 1 Maximum Wavelenght

|  |  |
| --- | --- |
| Comparison Agent | Λ Max |
| Gallic Acid | 756,8 nm |
| Quersetin | 435,5 nm |
| Tannic Acid | 743.5 nm |

**3.6 Preparation of Gallic Acid, quarsetin, and tannic acid Standar Curves**

              Standard curves were made with various concentrations of 20, 40, 60, 80, and 100 ppm measured at a maximum wavelength. The results are shown in table 2 .

**Table 2**

**Standard Concentration and Absorbance of Galic Acid, Quersetin, and Tannic Acid**

|  |  |  |  |
| --- | --- | --- | --- |
| **Concentration (ppm)** | **Absorbance of Gallic Acid** | **Absorbance of quersetin** | **Absorbance of tannic acid** |
| 20 | 0.0109 | 0.25 | 0.783 |
| 40 | 0.216 | 0.341 | 0.551 |
| 60 | 0.335 | 0.477 | 0.413 |
| 80 | 0.443 | 0.623 | 0.207 |
| 100 | 0.536 | 0.756 | 0.080 |

Table 3

Linear Regresion of Gallic Acid, Quersetin, And Taniic Acid Standard

|  |  |  |
| --- | --- | --- |
| Comparison Agent | Linear Regresion | Correlation coefficient (r) |
| Gallic Acid | y = 0.0054x + 0.0036 | 0.9986 |
| Quersetin | y = 0.0065x + 0.1012 | 0.9943 |
| Taniic Acid | y = 0.0087x-0.1175 | 0.991 |

Based on the table 3, it can be seen that correlation coefficient (r) of gallic acid, quersetin, and tannic acid the value of r approaches 1 which indicates that the regression equation is linear.

**3.7 Determination of Phenolic, Flavonoid, and Tannin Content**

**Table 4 Phenolic, Flavonoid, and Tannin Content of Kratom Stem Extract and Senggani Flower Extract**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
|  | Kratom Stem | | Senggani Flower | |
| Absorbance | Content (mg/gram of Extract) | Absorbance | Content (mg/gram of Extract) |
| Fenolic | 0,032 | 23.59 | 0,241 | 43.97 |
| Flavonoid | 0,130 | 10.65 | 0,028 | 11.26 |
| Tannin | 0,039 | 17.99 | 0,232 | 13.74 |

A total of 100 mg of kratom stem and senggani flowers ethanol extract were used as samples and three replications were made for data accuracy purposes. The absorbance data was entered into the linear regression that was obtained.

**V. Conclusions**

The total phenolic content in kratom stem extract was 23.59 mg / gram of extract, while in senggani flowers it was 43.97 mg / gram of extract. The total flavonoid level in kratom stem extract was 10.65 mg / gram of extract, while in senggani flowers it was 11.26 mg / gram of extract. The total tannin content in the kratom stem extract was 17.99 mg / gram of extract, while in the Senggani flower extract it was 13.74 mg / gram of extract.

The phenolic and flavonoid content in the ethanol extract of senggani flowers were greater than the ethanol extract of kratom stems. Meanwhile, the tannin content in the senggani flower extract was less than the ethanol extract of kratom stem.

**References**

1. Sahara M., Simanjuntak M., Aulia Y., Zai Y., Masdalena., 2019., **Uji Aktivitas Anti Diabetes Ekstrak Etanol Daun Senggani (*Melastoma Malabathrium* L) Pada Mencit Jantan Yang Diinduksi Aloksan**., Medan., Fakultas Kedokteran., Program Studi Pendidikan Dokter., Universitas Prima Indonesia.
2. Ridayani, Y., Andrie, M. dan Wijianto, B., 2013., **Uji Efek Sedatif Fraksi Etanol Daun Kratom (*Mitragyna speciosa* Korth.) pada Mencit Jantan Galur BALB/c**., IPI Jurnal Mahasiswa Fakultas Kedokteran UNTAN, 3, 1-9.
3. Alfian, Riza dan Susanti, H., 2012., **Penetapan kadar Fenolik Ekstrak Metanol Kelopak Bunga Rosella Merah (*Hibiscus sabdariffa* Linn.)dengan Variasi Tempat Tumbuh secara Spektrofotometri**., Jurnal Ilmiah Kefarmasian, 2(1), 73-80.
4. Wiendarlina Y. I., Rahminiwati M.,Gumelar T. F., 2018., **Aktivitas Hepatoprotektor Ekstrak Air Herba Pegagan Daun Kecil ( *Centella Asiatica L. Urb* ) Terhadap Tikus Putih Jantan *Sprague Dawley* L Yang Diinduksi Dengan Parasetamol**., Vol 8., No 1., Juni., Universitas Pakuan., Bogor.
5. Robinson, T., 1995., **Kandungan Organik Tumbuhan Tinggi**., Bandung: ITB P:71-73.
6. Ahmad A. R., Juwita., Ratulangi D. A. S.,Malik A., 2015., **Penetapan KadarFenolik Dan Flavonoid Total Ekstak Metanol Buah Dan Buan Patilaka ( *Etlingera Elatior (Jack)* R. M. SM )**., Vol 2., No 1., April., Universitas Muslim Indonesia., Makasar.
7. Apsari, Dwi, P., Susanti, H., 2011., **Penetapan Kadar Fenolik Total Ekstrak Metanol Kelopak Bunga Rosella Merah (Hibiscus Sabdariffa Linn) dengan Variasi Tempat Tumbuh secara Spektrofotometri**., Jurnal Ilmiah Kefarmasian, 2(1), 73-80.
8. Krisyanella., Susilawati N., Rivai H., 2013., **Pembuatan Dan Karakterisasi Serta Penentuan Kadar Flavanoid Dari Ekstrak Kering Herba Meniran ( *Phyllanthus Niruri* L . )**.,Jurnal Farmasi Higea., Vol 5., No 1., Universitas Andalas., Padang.
9. Karismawati M., Utami P. S., Wahyuningum R.., 2009., **Penetapan Kadar Tanin Dalam Infusa Daun Salam (Syzygium Polyanthum(Wight.) Walp)) Secara Spektrofotometri Sinar Tampak., Vol.06., No. 01**., April., Universitas Muhammadiyah., Purwokerto.
10. Mukhriani, Nonnci F. Y., Mumang., 2014., **Penetapan Kadar Tanin Total Ekstrak Biji Jintan Hitam ( *Nigella Sativa )* Secara Spektrofotometri UV – Vis**., Vol. 2., No. 4., Universitas Islam Negeri Alauddin., Makassar.
11. Agoes., G., 2007., **Teknologi Bahan Alam**, ITB Press Bandung.
12. Harbone, J.R., 1987., **Metode Fitokimia Penuntun Cara Modern Mengekstraksi Tumbuhan**., Terjemahan ITB. Bandung. Padmawinata Edisi kedua.